18th International Conference on Retinal Proteins

Computational **Models for Rhodopsins:** from Primary Structures to Optical Properties Mikhail N. Ryazantsev

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Any computational model is an approximation.

All computational models have to be validated against experimental data.



Computational models have to provide insight into a problem, not only numbers.





There are several thousands unique rhodopsins with amino acid sequence available.



There are only (March 2018) unique X-ray rhodopsin structures in the **RCSB** database.









HOW TO PREDICT **A PROTEIN STRUCTUR** TO VALIDATE IT?

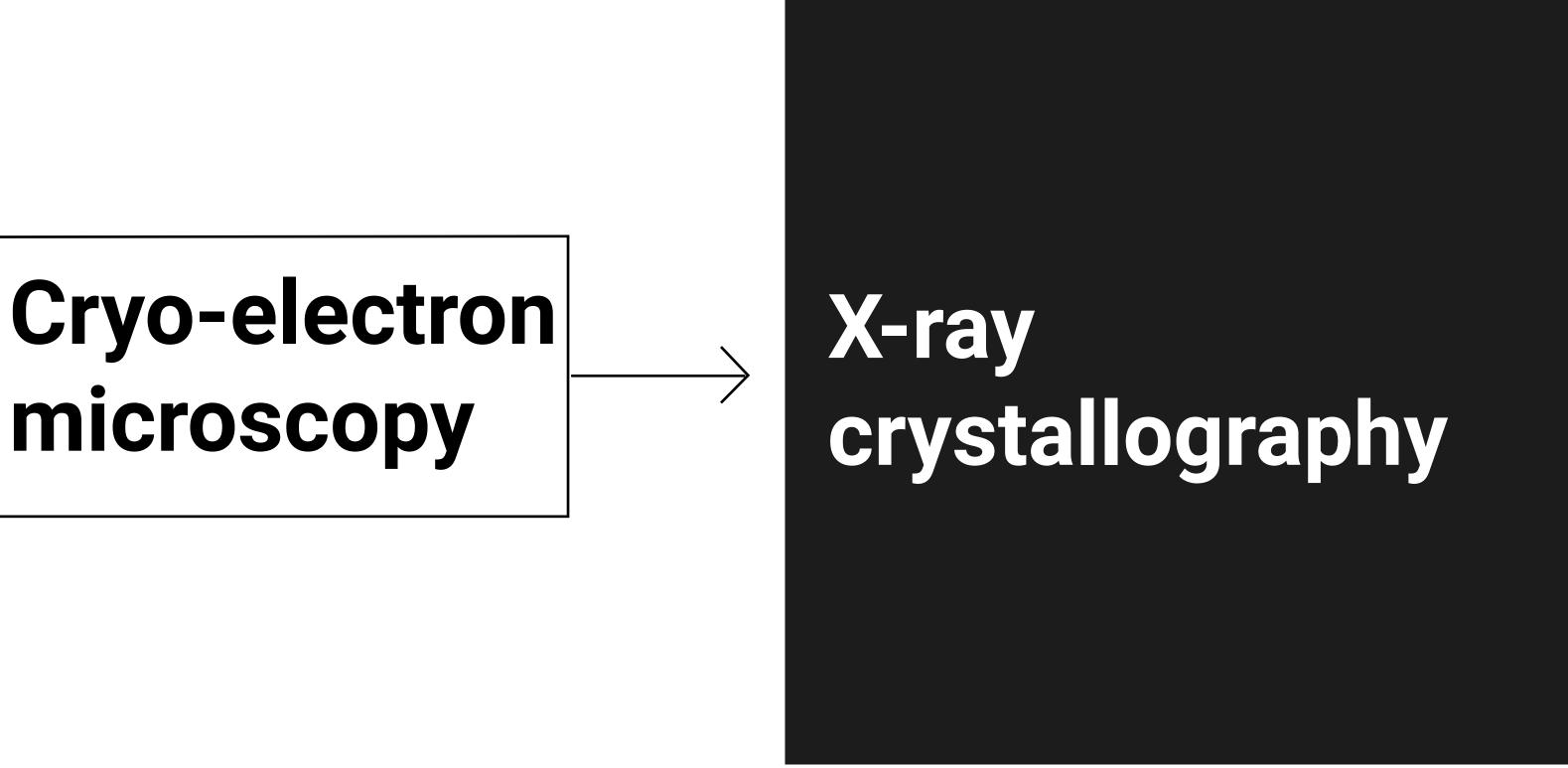


DECREASE OF CONFORMATIONAL SPACE FOR STRUCTURE PREDICTION

PureHomologyCryo-ab initiomodelsmicro

How to validate predicted model:





Against its X-ray structure (compare the geometries) Against experimental optical properties.



VALIDATION OF PREDICTED MODEL **AGAINST PROTEIN X-RAY STRUCTURE**

To unambiguously evaluate the quality of homology models we produced models only for rhodopsins whose struture has been determined experimentally.

In this way, we could compare the resulting model with the corresponding experimental structure using common metrics (RMSD, GDT-HA).

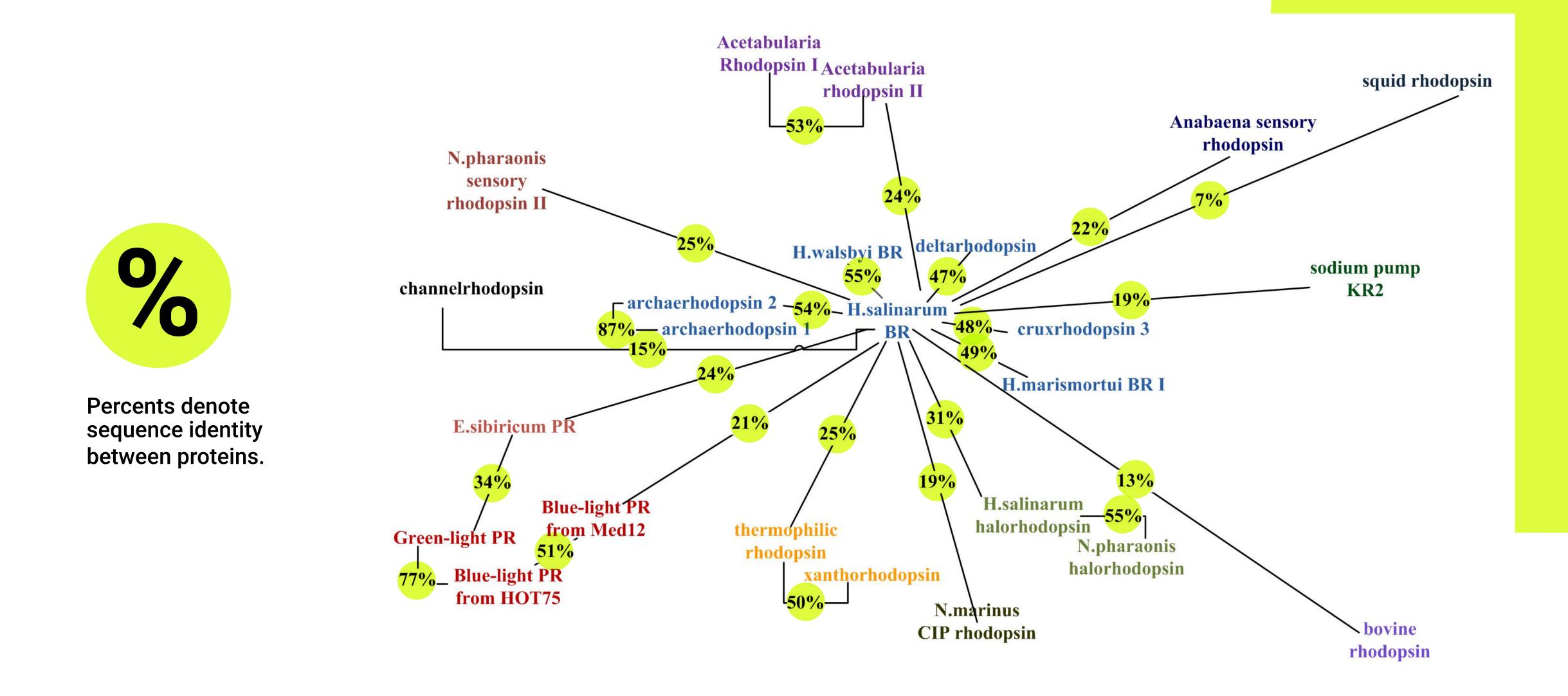
In our study we formed pairs of experimental structures (A and B), and predicted structure of A using structure of B as a template and vice versa.







CLUSTERING OF RHODOPSINS WITH AVAILABLE X-RAY STRUCTURE





7

PREDICTION OF RHODOPSIN THREE-DIMENSIONAL STRUCTURES

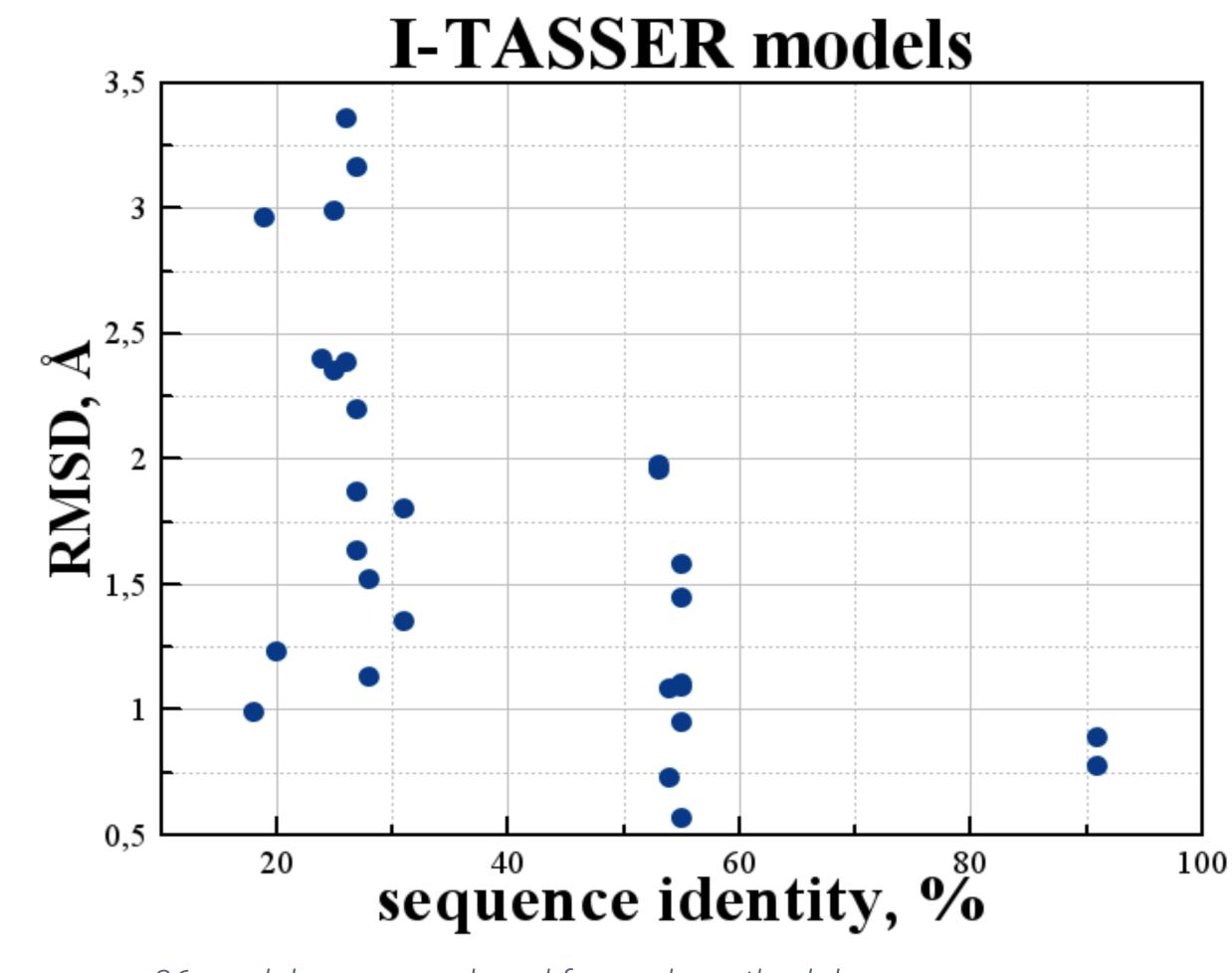
Alignment:	Structure building:
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MP-T AlignMe MUSTER Medeller I-TASSER RosettaCM

Ca-RMSD:

Root-Mean-Square Deviation of corresponding Ca atoms after structural superposition of model onto experimental structure.



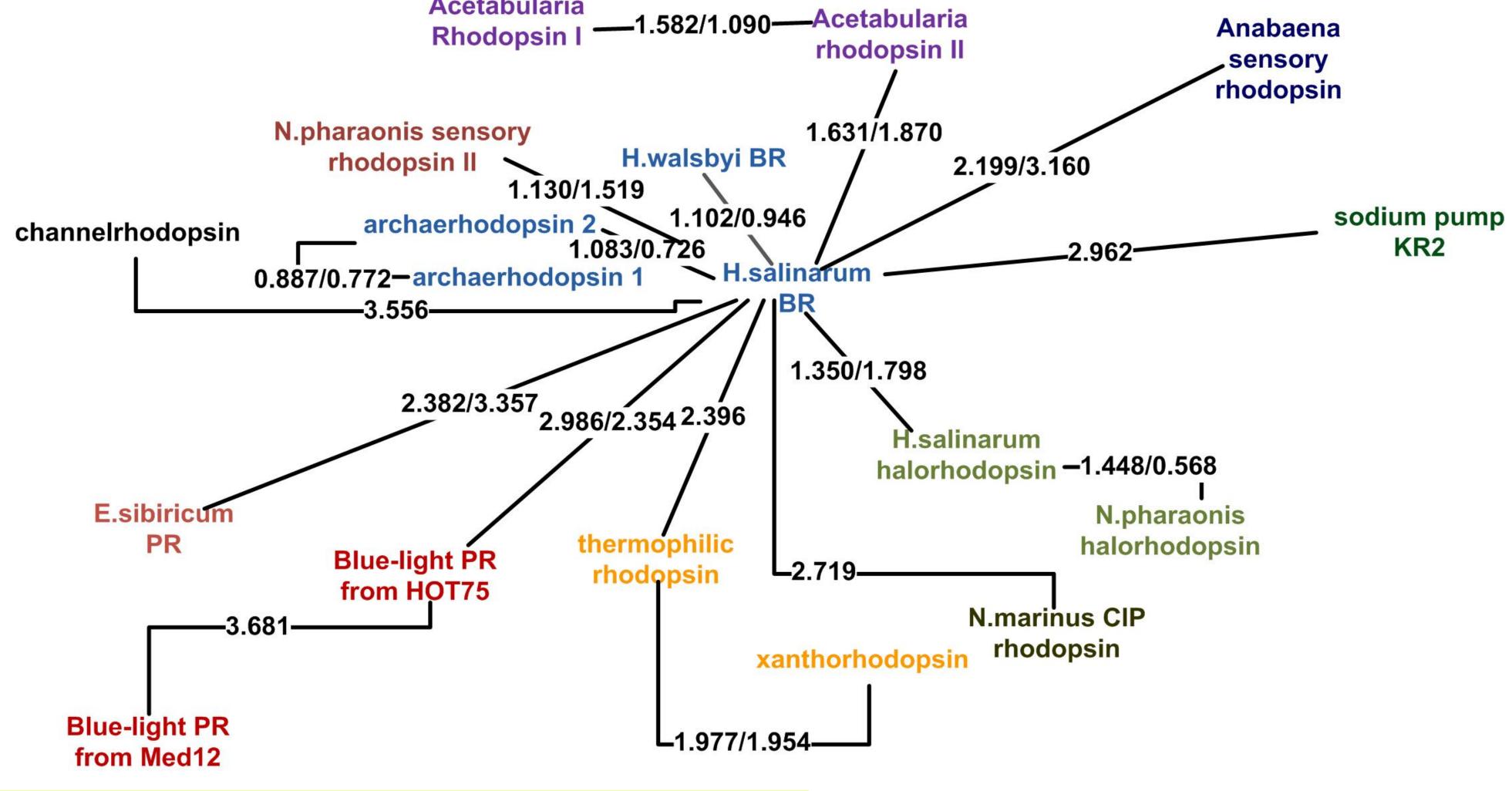


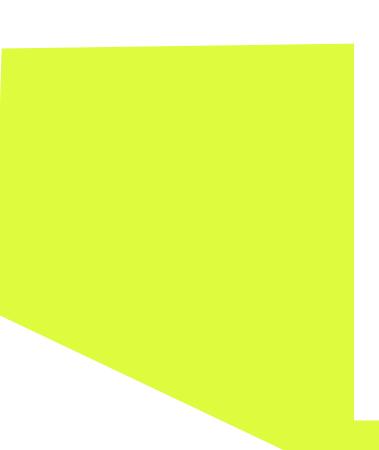
36 models were produced for each methodology (alignment + structure building)



CLUSTERING BASED ON PREDICTED MODEL QUALITY

Acetabularia

















We obtained models with:

Average intracluster **Ca-RMSD** less than 1.5 Å

Average overall Cα-RMSD: around 2 Å

For the transmembrane part of rhodopsins average **RMSD** is less than **1** Å

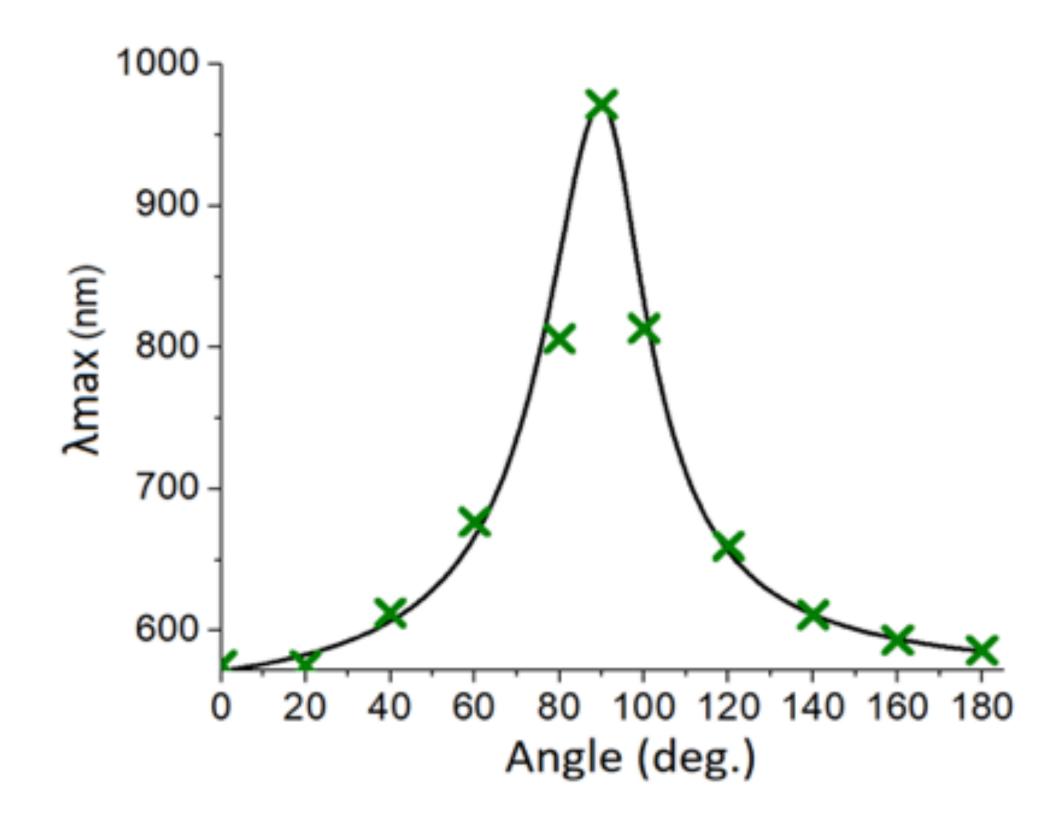
Nikolaev, D.M., Shtyrov, A.A., Panov, M.S., Jamal, A., Chakchir, O.B., Kochemirovsky, V.A., Olivucci, M. and Ryazantsev, M.N., 2018. A Comparative Study of Modern Homology Modeling Algorithms for Rhodopsin Structure Prediction. ACS omega, 3(7), pp.7555-7566.



Conclusion: with the right choice of template and methodology it is possible to predict rhodopsins with quality close to the X-ray one

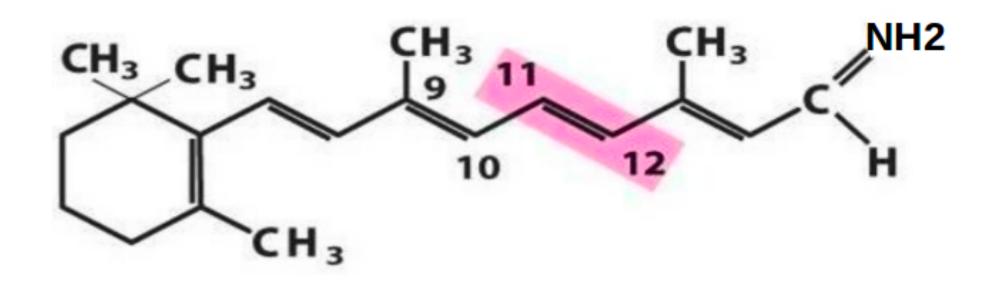


Influence of retinal distortion: rotation of C11=C12 bond

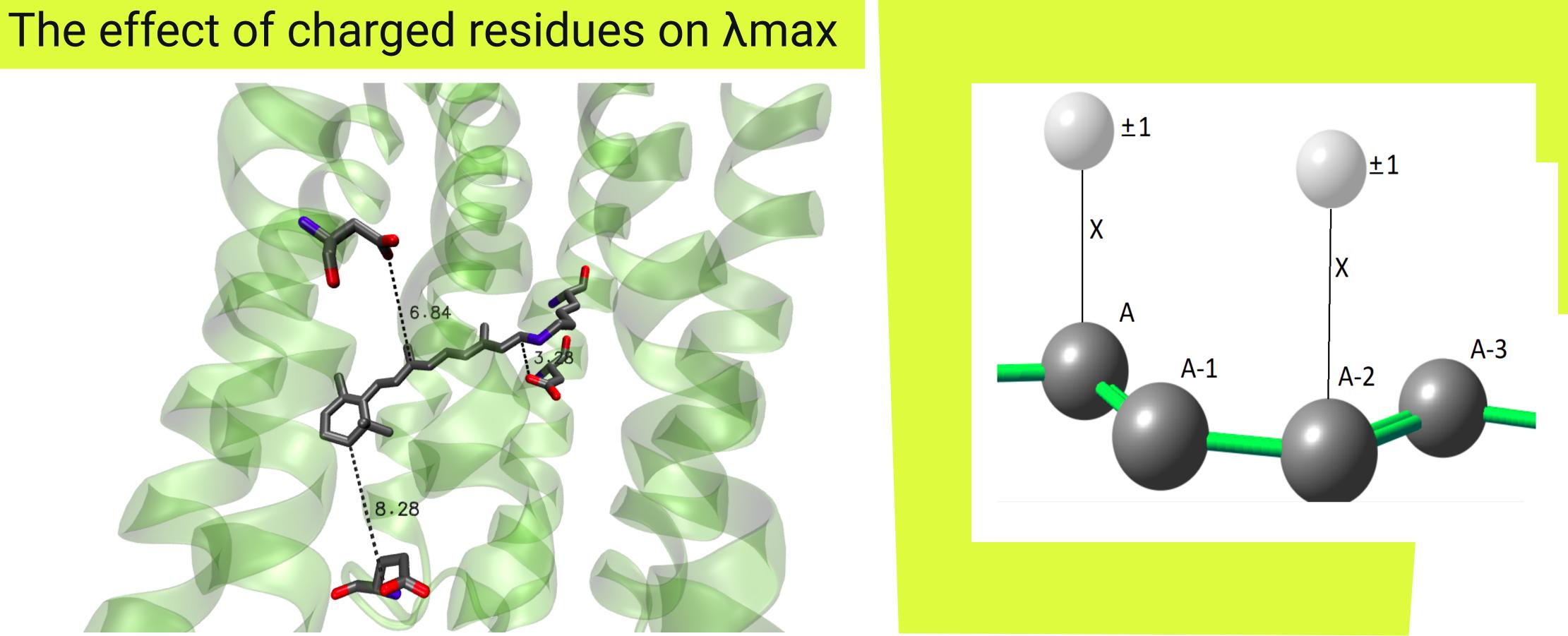








SORCI+Q/ 6-31g*//B3lyp/6-31g*

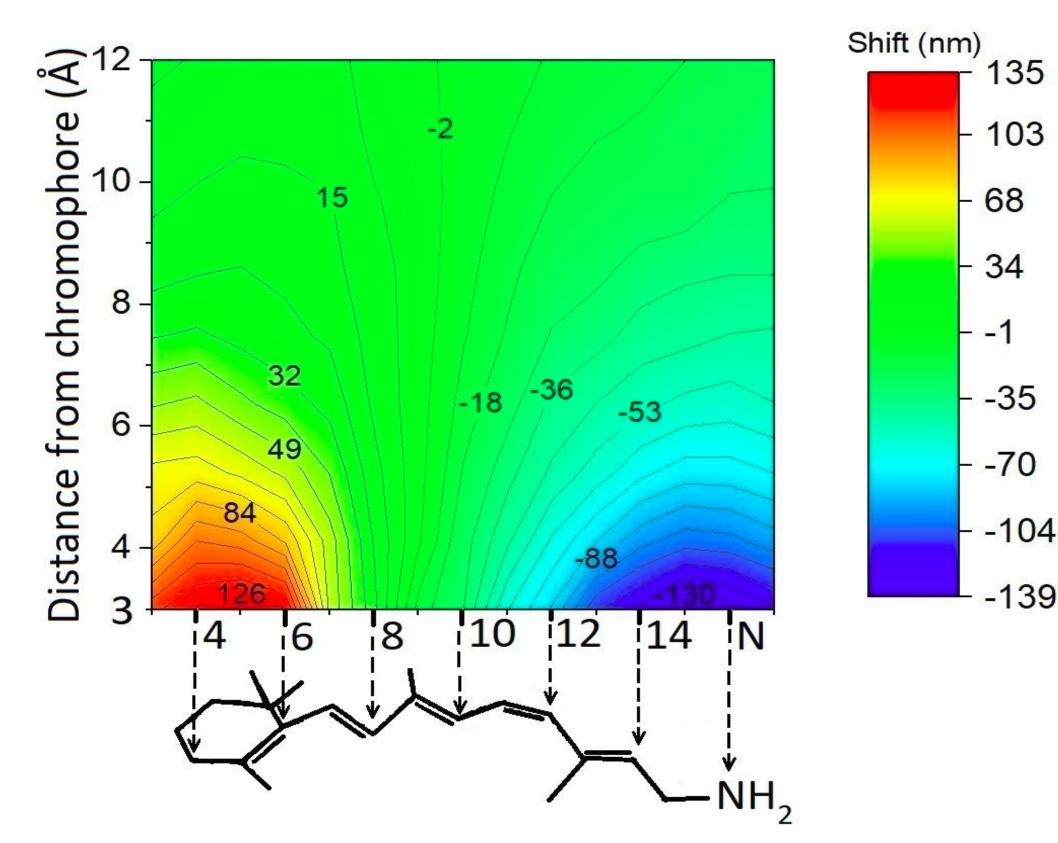








The effect of point negative charges on λ max







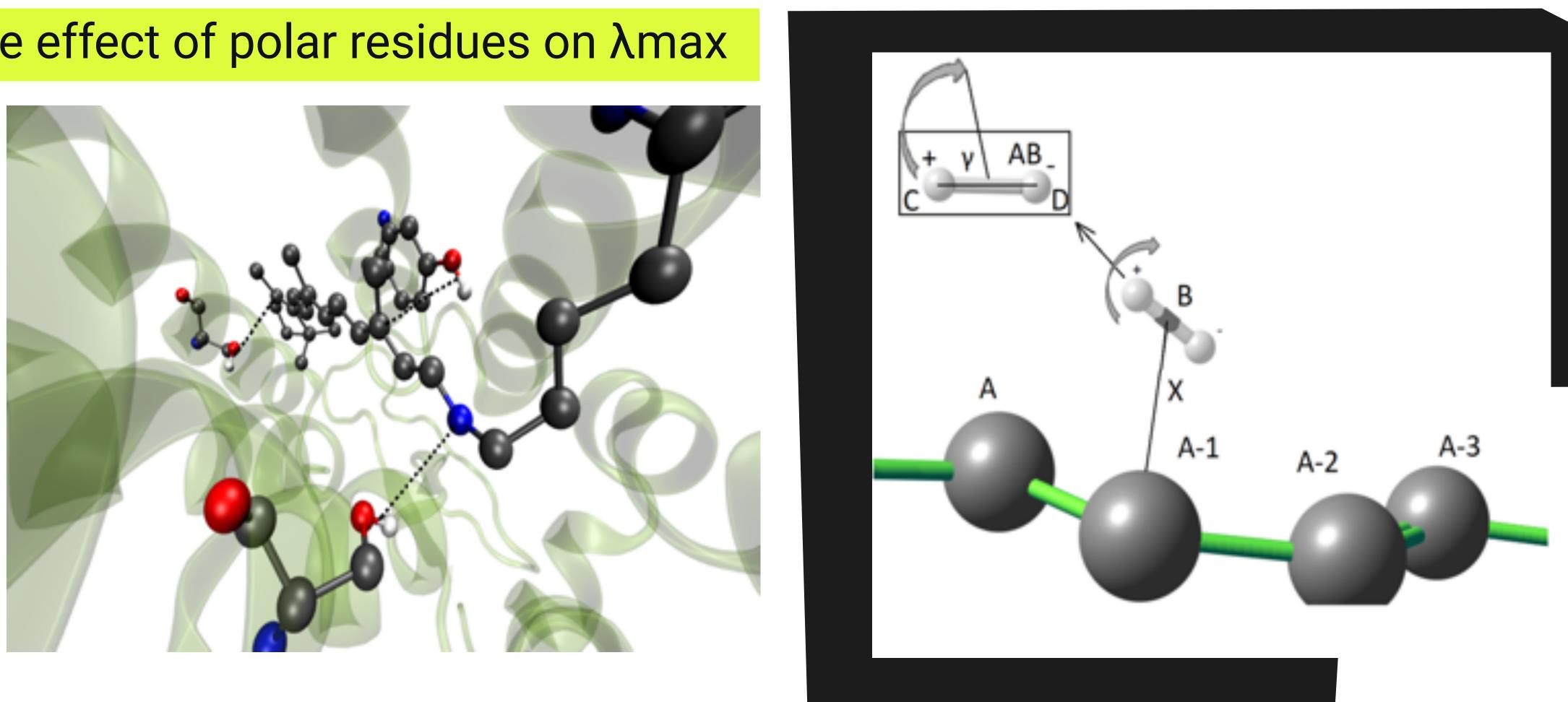
- 135
- 103
- -70
- -139

SORCI+Q/ 6-31g*//B3lyp/6-31g*



13

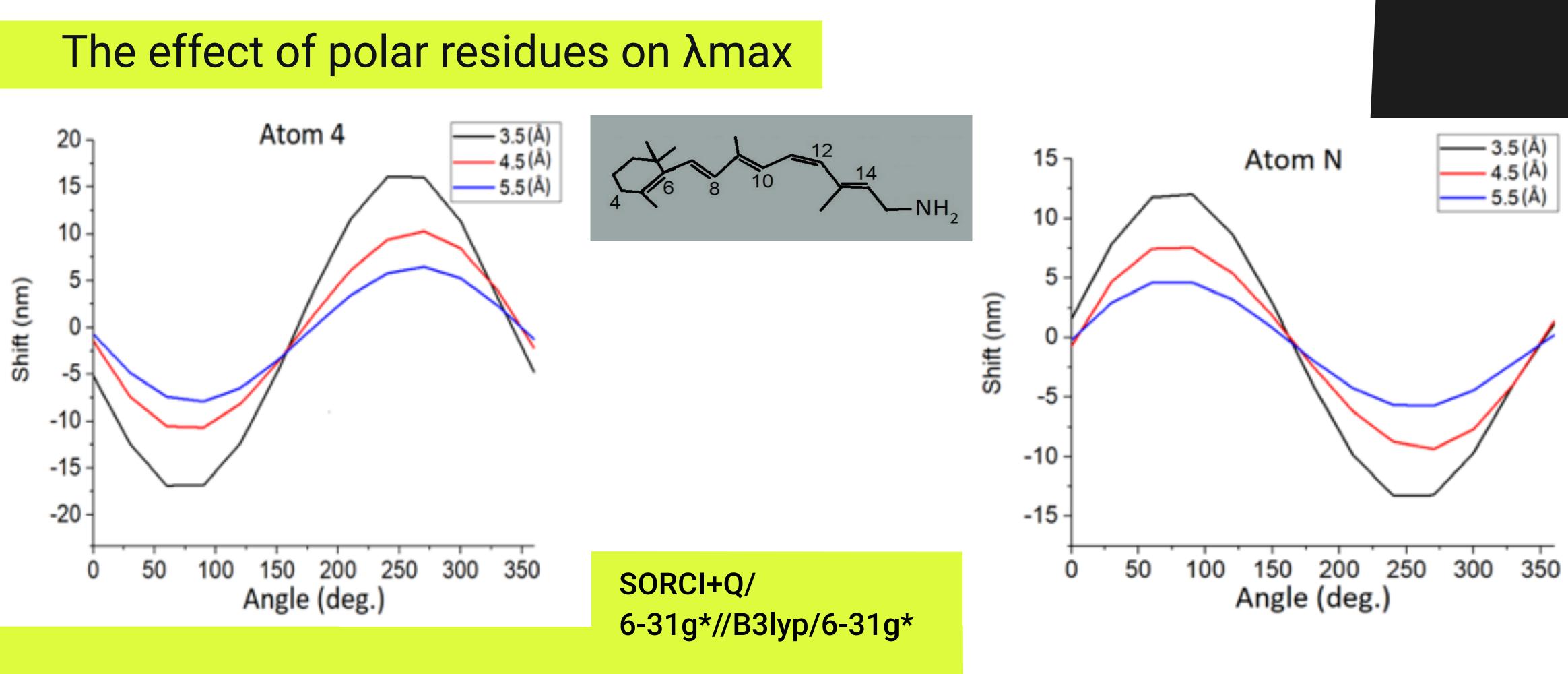
The effect of polar residues on λmax







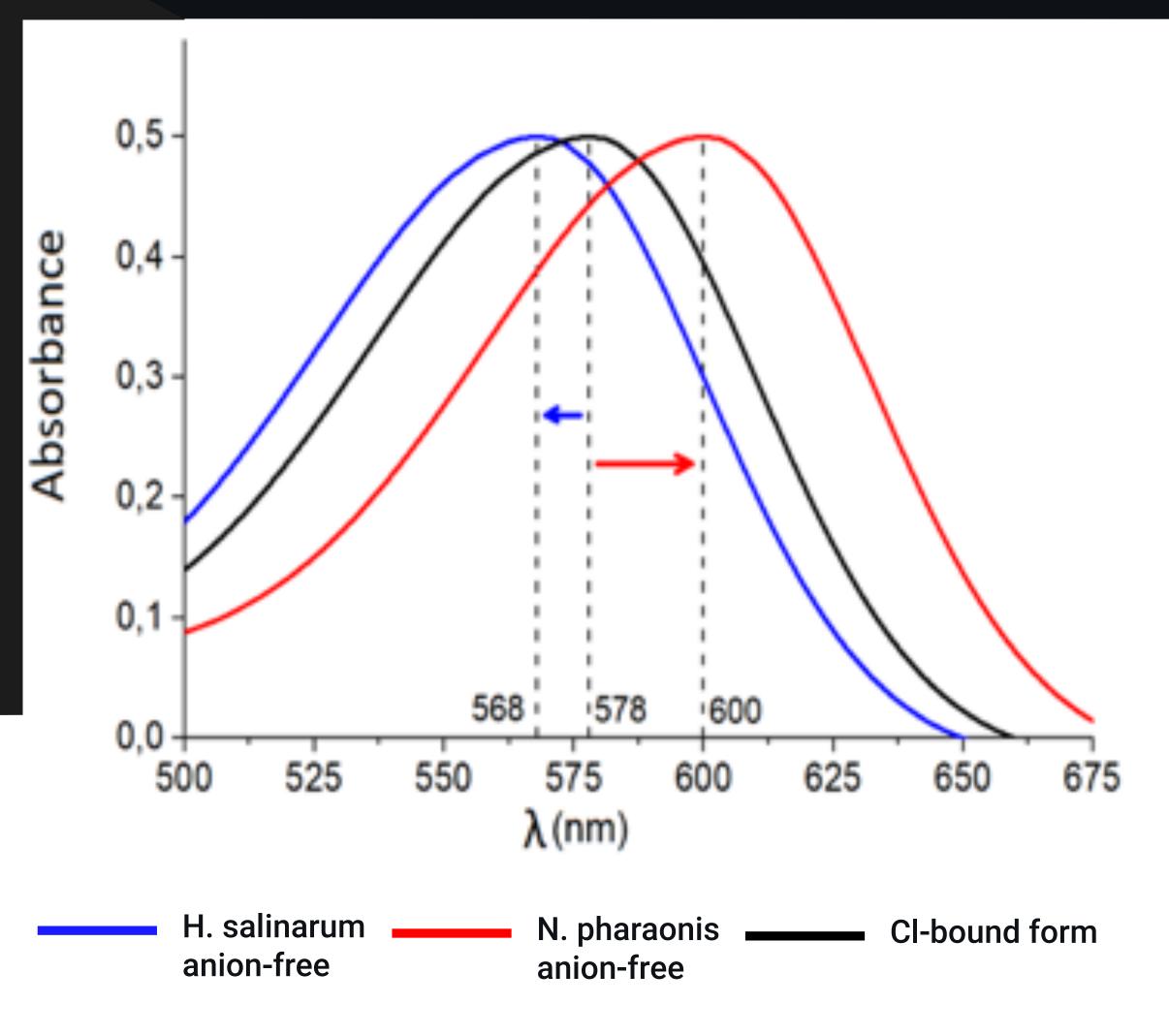
14







CASE STUDY: CL-INDUCED SPECTRAL SHIFT IN HALORHODOPSINS



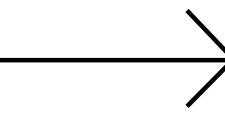
Ryazantsev, M.N., Altun, A. and Morokuma, K., 2012. Color tuning in rhodopsins: the origin of the spectral shift between the chloride-bound and anion-free forms of halorhodopsin. **JACS, 134(12)**.

Cl-bound spectral shift:

	Calculated	Experimer
N. Pharaonis	+ 29	+ 22
H. Salinarum	- 10	- 9











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Nikolaev D. **Dogonasheva O.** Panov M. Shtyrov A. **Belyaev L**. Lebedeva M. **Rusanov V.** Derevyankina G.

Fomin A. Kravtsov D. Blokhin N. Osipov D. Firsov M. Boitsov V. Belyaeva E











